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Copyright Holder	Springer Nature Switzerland AG	
Corresponding Author	Family Name	<b>Maqbool</b>
	Particle	
	Given Name	<b>Farhana</b>
	Suffix	
	Division	Department of Microbiology
	Organization/University	Hazara University
	Address	Mansehra, KPK, Pakistan
	Email	drfarhana@hu.edu.pk
Author	Family Name	<b>Bhatti</b>
	Particle	
	Given Name	<b>Zulfiqar A.</b>
	Suffix	
	Division	Department of Environmental Science
	Organization/University	COMSATS University Islamabad, Abbottabad Campus
	Address	Abbottabad, KPK, Pakistan
	Author	Family Name
Particle		
Given Name		<b>Muhammad Faisal</b>
Suffix		
Division		Department of Microbiology
Organization/University		Hazara University
Address		Mansehra, KPK, Pakistan
Author		Family Name
	Particle	
	Given Name	<b>Ibrar</b>
	Suffix	
	Division	Department of Microbiology
	Organization/University	Abbottabad University of Science and Technology
	Address	Abbottabad, KP, Pakistan

Author	Family Name	<b>Zhao</b>
	Particle	
	Given Name	<b>Yang Guo</b>
	Suffix	
	Division	College of Environmental Science and Engineering
	Organization/University	Ocean University of China
	Address	Qingdao, China
Author	Family Name	<b>Sajid</b>
	Particle	
	Given Name	<b>Muhammad</b>
	Suffix	
	Division	Department of Biochemistry
	Organization/University	Hazara University
	Address	Mansehra, KPK, Pakistan
Author	Family Name	<b>Umm-e-Kalsoom</b>
	Particle	
	Given Name	
	Suffix	
	Division	Department of Biochemistry
	Organization/University	Hazara University
	Address	Mansehra, KPK, Pakistan
Author	Family Name	<b>Mehmood</b>
	Particle	
	Given Name	<b>Qaiser</b>
	Suffix	
	Division	Department of Environmental Science
	Organization/University	COMSATS University Islamabad, Abbottabad Campus
	Address	Abbottabad, KPK, Pakistan
Author	Family Name	<b>Nawaz</b>
	Particle	
	Given Name	<b>Faiza</b>
	Suffix	

Division	Department of Microbiology
Organization/University	Hazara University
Address	Mansehra, KPK, Pakistan

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Abstract	Breakdown of the crude oil contaminant by soil microbial community in the bioremediation can be enhanced in the presence of plant rhizosphere, but plant cannot grow and tolerate the high concentration of hydrocarbon. Composting of the crude oil-contaminated soil was found to be an effective technology to decrease the pollutant concentration to the acceptable level for plant growth. Phytoremediation with native plants followed with composting can be performed well, but the microbial abundance in the contaminated soil usually becomes much low as time passes. In order to enhance the degradation potential of heterotrophic microorganisms in bioremediation, different bioaugmentation techniques are used, while few researches also used plant rhizosphere along with it, but the results are ambiguous. The present chapter will explore this ambiguity by providing the complete description about the bioremediation and the facts that plant rhizosphere selects microbial community, which is more suitable for pollutant degradation than addition of microbial culture in soil. Rhizodegradation followed with composting is found to be a useful treatment method for extremely polluted soil, and bioaugmentation with immobilized and free bacterial culture was found more effective without plant.
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# Chapter 7 1

## Inoculum Addition in the Presence 2

### of Plant Rhizosphere for Petroleum- 3

### Polluted Soil Remediation 4

**Farhana Maqbool, Zulfiqar A. Bhatti, Muhammad Faisal Siddiqui, 5**  
**Ibrar Khan, Yang Guo Zhao, Muhammad Sajid, Umm-e-Kalsoom, 6**  
**Qaiser Mehmood, and Faiza Nawaz 7**

## 7.1 Introduction 8

Composting of contaminated soil is one of the feasible methods to clean up petro- 9 [AUI](#)  
leum hydrocarbon in the biopiles by the addition of nutrients and amendments, 10  
which enhances the hydrocarbon degradation and improve soil quality (Van 11  
Gestel et al. 2003). However, the contents of the petroleum hydrocarbons and 12  
their decomposed matter present in the soil after composting are often above envi- 13  
ronmental standards (Zhang et al. 2010). Bioremediation is the conversion of 14  
complex organic contaminant into simpler inorganic compounds including car- 15  
bon dioxide, water, and cell biomass by biological agents like microorganisms 16  
(Das and Chandran 2011). Due to the presence of pollutant in the soil, microor- 17  
ganism including bacteria, fungi, and yeast that prefer the chemicals as a source 18  
of food and energy can be flourished. In the soil, mineralization rate contaminant 19  
depends on the microbial activity and abundance in the soils. Pollutant degrada- 20  
tion is extremely dependent on the presence of electron provider or acceptors, 21

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F. Maqbool (✉) · M. F. Siddiqui · F. Nawaz  
Department of Microbiology, Hazara University, Mansehra, KPK, Pakistan  
e-mail: [drfarhana@hu.edu.pk](mailto:drfarhana@hu.edu.pk)

Z. A. Bhatti · Q. Mehmood  
Department of Environmental Science, COMSATS University Islamabad, Abbottabad  
Campus, Abbottabad, KPK, Pakistan

I. Khan  
Department of Microbiology, Abbottabad University of Science and Technology,  
Abbottabad, KP, Pakistan

Y. G. Zhao  
College of Environmental Science and Engineering, Ocean University of China,  
Qingdao, China

M. Sajid · Umm-e-Kalsoom  
Department of Biochemistry, Hazara University, Mansehra, KPK, Pakistan

22 presence of co-metabolites, absence of inorganic nutrients, plant vitamins and  
 23 hormones, as well as microbial competition (ITRC 2009).

24 Bioremediation methods, which enhance the biodegradation rates, include  
 25 natural attenuation without any interference, use of seed culture of hydrocar-  
 26 bon degraders, biostimulation with nutrient addition for improved microbial  
 27 growth, and rhizospheric biodegradation (Odokuma and Dickson 2003;  
 28 Bhatti et al. 2017). Microbes spread in the rhizosphere, and their number  
 29 increases due to the availability of nutrients which are released by plants  
 30 including C, N, S, and P. Few studies used bioaugmentation in the presence  
 31 of plant to remediate the petroleum-polluted soil (Kirk et al. 2005; Baek  
 32 et al. 2004).

33 Through bioaugmentation desired microbes and their enzymes increase,  
 34 but this process does not work always due to the inability of culture to adjust  
 35 in new environment and to compete with native microbes (Gerardi 2016;  
 36 Ramos et al. 2010). Microbial entrapment in solid media is used for the pro-  
 37 tection and survival of inoculated culture, and the most effective microbial  
 38 immobilization was cell entrapment into a porous matter (Partovinia and  
 39 Rasekh 2018). The immobilized cells have proved stability under different  
 40 environmental conditions for petroleum-contaminated soil (Gao et al. 2011).  
 41 The present chapter reveals different strategies of complete biodegradation of  
 42 highly petroleum-polluted soil, limitation, and outcomes of these techniques.

## 43 7.2 Different Strategies of Bioremediation

### 44 7.2.1 Composting of Highly Petroleum-Polluted Soil

45 Composting is the addition of biomaterial and biogenic bulking agents  
 46 used for aeration and to adjust water, nutrient, and pH of the contaminated  
 47 soil (Kästner and Miltner 2016). Composting of highly petroleum-contam-  
 48 inated soil is a very encouraging technique, and it can be performed at the  
 49 first step before phytoremediation to lower down the concentration to  
 50 acceptable level for plant growth. Maize straw, pine wood chips, and soy-  
 51 bean cake were used as bulking agent in composting to lower down the  
 52 total petroleum hydrocarbon (TPH) concentration from 17,900 to  
 53 3700 mg kg<sup>-1</sup> (Wang et al. 2011). This composted soil was further used to  
 54 lower down the concentration up to 673 mg kg<sup>-1</sup> using *Sesbania canna-*  
 55 *bina*, a coastal halophyte (Table 7.1 shows all results of the project)  
 56 (Farhana et al. 2012). Jørgensen et al. (2000) used bark chips as the bulk-  
 57 ing agent with inoculum to compost the lubricating oil-contaminated soil;  
 58 70% removal rate was observed with organic matter addition and no effect  
 59 of inoculum addition. Another study used cow bed and potato peelings for  
 60 high asphaltenic fuel oil composting, and the result showed that with

**Table 7.1** Different bioremediation approaches for the Shengli Oil Field petroleum-polluted soil and their maximum removal percentages

S. no.	Technique	Maximum bioremediation duration (days)	Removal percentage of petroleum hydrocarbon (%)
1	Bioaugmentation with free culture inoculation	70	39
2	Bioaugmentation with immobilized culture inoculation	70	47
3	Addition of compost in contaminated soil	90	45
4	Phytoremediation with seepweed	90	42
5	Phytoremediation with <i>S. cannabina</i>	90	73
6	<i>S. cannabina</i> in the presence of free culture inoculation	120	73
7	<i>S. cannabina</i> in the presence of immobilized culture inoculation	120	68

<sup>1,2</sup>Wang et al. (2012), <sup>3,4</sup>(Wang et al. (2011), <sup>5,6,7</sup>Farhana et al. (2012)

autochthonous bacteria, fragmentation into easily degradable smaller structure took place (Martin-Gil et al. 2008).

## 7.2.2 Enhanced Degradation of Pollutants with Bioaugmentation

Sometimes biodegradation fails due to insufficient indigenous oil degrader population or their incapability of degrading the broad range of toxic fractions present in the environment (Hussein 2006). In this situation, bioaugmentation was found to be a promising low-cost technique in which effective bacterial culture or microbial community capable of degrading hydrocarbons is added to the contaminated soil (Zawierucha and Malina 2011). Bioaugmentation can be achieved in three ways, first, autochthonous bioaugmentation in which seed cultures are obtained from the original contaminated site, re-cultured in the lab, and reused in the same soil which needs treatment (Ueno et al. 2007). The second method is that seed culture is taken from one site and used in different contaminated site, and the third method is the use of genetically modified organism with degradative potential (Vogel and Walter 2001). Among these autochthonous bioaugmentation is of great interest because it is found to be best adopted with the contaminated environment.

Many researchers used bioaugmentation alone for crude oil degradation (Bento et al. 2005; Yu et al. 2005), but few researches are found in the field of rhizosphere bioaugmentation (Peng et al. 2009; Ma et al. 2010; Cai et al. 2010; Tang et al. 2010; Thangarajan et al. 2011) (Table 7.2). The use of higher plants to enhance

83 removal of contaminants from soil although low cost than other remediation tech-  
84 niques often does not result in complete removal of contaminants (Banks et al.  
85 2003); therefore some researchers tried to use inoculum addition in the presence  
86 of plant. Bioaugmentation required successful and safe introduction of microbial  
87 cells into uncontained environments (Trevors et al. 1993).

88 Ramos et al. (2010) explained that inoculated cultures sometimes fail to  
89 resist and survive in contaminated soil or in their native living place soil.  
90 Seed culture should be highly stable and resistant to different environmen-  
91 tal conditions (Burken 2004). These obstacles can be overcome using some  
92 suitable inoculation techniques.

### 93 7.2.2.1 Liquid Microbial Culture Inoculation

94 In previous studies for bioaugmentation in soil, broth culture inoculation  
95 such as free culture spraying procedures or culture in semisolid media, i.e.,  
96 immobilization techniques, was used (Trevors et al. 1994). The use of inoc-  
97 ulated culture is not always acceptable, since inoculation experiments have  
98 shown ambiguous results in comparison to degradation by native microbial  
99 population (Leahy and Colwell 1990). Kastner et al. (1998) used several  
100 strains of PAH-degrading bacteria, but after introduction into artificially  
101 PAH-contaminated soil, no degradation was observed; further experiment  
102 suggested that inoculation with mineral salt medium inhibited bacterial sur-  
103 vival and growth as well as their degradation activity; while using water  
104 without increasing salinity, no inhibition of autochthonous bacterial strain  
105 took place (Hosokawa et al. 2009).

### 106 7.2.2.2 Inoculation with Immobilized Microbial Culture

107 Captivity of enzyme or whole bacterial cell inside or outside the carrier matrix  
108 is known as immobilization (Partovinia and Rasekh 2018). For petroleum  
109 hydrocarbon biodegradation, microbial attachment on solid surface, entrapment  
110 in a gel or membrane, or encapsulation into carrier was used (Hussein 2006).  
111 Microbial culture for bioaugmentation in an immobilized form may offer more  
112 complete and more rapid degradation, which is easy to handle, can be reuti-  
113 lized, and has high tolerance to pH and temperature changes (Partovinia and  
114 Rasekh 2018). Immobilized matrices provide sustained microbial population by  
115 continuous release of bacteria in the soil, water, and sediments. Although in  
116 many studies culture used in immobilized beads form, still most of the studies  
117 have not addressed the growth dynamics of different bacterial strains inside the  
118 beads (Bazot and Lebeau 2009).

119 There are two types of hydrogel material for cell immobilization, natu-  
120 ral and synthetic. Natural material includes agar, agarose, polyacryl-  
121 amides, carrageenan, and alginate (Leenen et al. 1996) (Table 7.3).



**Table 7.2** Researches on bioaugmentation in bioremediation of petroleum-contaminated soil

S. no.	Strategies	Pollutants	Outcome	References
1	Natural attenuation	TPH	Effective than biostimulation	(Mishra et al. (2001))
2	Phytoremediation (grasses)	Diesel and heavy oil	Noneffective in old contaminated soil	Banks et al. (2003)
3	Bioaugmentation	TPH	Effective than NA*/B <sup>a</sup>	Bento et al. (2005)
4	Bioaugmentation	PAH	Noneffective than NA*	Yu et al. (2005)
5	Bioaugmentation	PAH	Noneffective	Herwijnen et al. (2006)
6	Phytoremediation	TPH	Effective than NA*	Peng et al. (2009)
7	Phytoremediation	PAH	Effective in freshly contaminated soil than old	Ma et al. (2010)
8	Phytoremediation ( <i>Impatiens balsamina L</i> )	TPH	Effective than natural degradation	Cai et al. (2010)
9	Phytoremediation and bioaugmentation	Petroleum	Combine treatment effective than single	Tang et al. (2010)
10	Bioaugmentation	TPH	Noneffective than NA*/B <sup>a</sup>	Thangarajan et al. (2011)
11	Bioaugmentation	Aromatic and asphaltic	Effective than indigenous	Gonzalez et al. (2005)
12	Bioaugmentation	Crude oil	Mix culture effective than single spp.	Rahman et al. (2002)

\*NA natural attenuation, B<sup>a</sup> bioaugmentation

Alginate is used widespread due to its nontoxic and nutritional behavior for bacterial cell immobilization. The incorporation of some adsorbents into alginate beads is used to transport pollutants inside and outside of the beads (Zhang et al. 2008). Sodium alginate matrix has hindrance to mass and air transfer inside the dense gel layer (Mikkelsen and Elgsaeter 1995). Diatomite was incorporated to overcome this problem (Farhana et al. 2012). Diatomite (SiO<sub>2</sub> nH<sub>2</sub>O) is off-white to white color, soft, light in weight sedimentary rock composed mainly of silica microfossils of aquatic algae. It is highly porous with good sorption ability, is nonreactive with other chemicals, and has low density and large surface area; due to these unique properties, this material is used in cell immobilization for hydrocarbon degradation (Nenadovic et al. 2009). Wang et al. (2012) in their study found that after 20 days of oil degrader inoculation in the soil, the maximum degradation rate in the sodium alginate diatomite (SAD)-immobilized systems reached up to 29.8%, significantly higher ( $P < 0.05$ ) than free cells (21.1%). Moreover, both microbial number and total microbial activity reached to highly significant level ( $P < 0.05$ ) in the immobi-

139 lized culture than free culture inoculation systems at a same initial  
140 inoculation amount. In the presence of plant rhizosphere, this immobi-  
141 lized inoculum was ineffective (Table 7.1) (Farhana et al. 2012) because  
142 sodium alginate diatomite carrier has short life and degraded within 20  
143 days and plant growth takes more time (Mikkelsen and Elgsaeter 1995).

### 144 7.2.3 *Rhizoremediation of Pollutants*

145 Pollutant breakdown in the rhizosphere is improved due to the increase in the  
146 microbial activity than non-rhizosphere; this process is generally known as phy-  
147 tostimulation or plant assisted microbial degradation (Donnelly et al. 1994).  
148 Rhizodegradation involves the immobilization and removal of the pollutants  
149 which is strongly dependent on the rhizospheric processes including enhance-  
150 ment of microbial degradation. The number of PAH degraders and total microbial  
151 count were found higher in the rhizospheric region of ryegrass and clover than in  
152 the un-vegetated soil (Ma et al. 2010).

153 Symbiotic association is observed around the plant rhizosphere and their asso-  
154 ciated microbes due to the release of root exudates which provide necessary nutri-  
155 ents for the survival of microbes and in turn microbes responsible for improved  
156 soil conditions for plant growth. Especially, plants help in softening of soil due to  
157 aeration and water transport into the rhizosphere (Fig. 7.1). Plants also release  
158 allelopathic agents which suppress the growth of other plants in the same soil and  
159 protect them from competition, soil pathogens, toxins, and chemicals released  
160 from the unwanted plants.

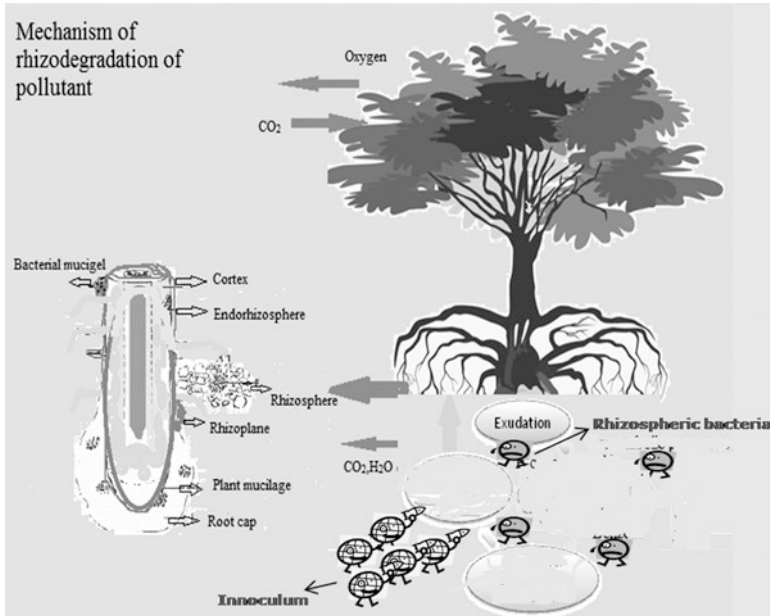
161 In specific plant-microbe interaction, plant detects toxicants and secretes spe-  
162 cific compounds in response to that stimuli and subsequently promotes the growth  
163 and activity of specific bacterial community (Fang et al. 2001). Due to this reason  
164 sometimes inhibition of inoculated bacterial culture was observed which inter-  
165 feres with the natural plant-microbe interaction and leads to the failure in enhanced  
166 degradation by bioaugmentation (Farhana et al. 2012) (Fig. 7.1). Nonspecific  
167 interaction occurs when the contaminants themselves may have similarities to the  
168 phytochemical which are released by the plant naturally (Phillips et al. 2012).  
169 These phytochemicals are used by the rhizospheric microbes as a primary carbon  
170 source, and co-metabolic process starts which slows down the pollutant degrada-  
171 tion. Sometimes pollutants' degradative enzymes produced by microbes can also  
172 be produced by the plant itself. These secondary plant metabolites (SPMEs) are  
173 very diverse in nature, having structural similarities with organic pollutant which  
174 play an important role in the production of many degradative enzymes (Jha et al.  
175 2015). Examples of analogy included PAH, their structures are similar to the root  
176 exudates morin (Donnelly et al. 1994), and pyrene resembles SPME confusarine  
177 released by plants (Singer et al. 2003). The degradation capacity increased in the  
178 presence of exudates acetate and alanine while decreased when malonate was

**Table 7.3** Summary of the researches on different immobilization strategies for bioaugmentation t3.1

Material for immobilization	Target pollutant	Microorganisms type	Effect	Reference	t3.2
Biofix and Drizit	Petrol	<i>Pseudomonas fluorescens</i>	Best results than	Wilson and Bradley (1996)	t3.3 t3.4 t3.5
Sodium alginate	Phenol, ortho- and Para-cresol	Methanogenic consortium	Twofold higher than free culture	Guiot et al. (2000)	t3.6 t3.7
Polyvinyl alcohol and Drizit	Diesel	Mix hydrocarbon degraders	Best results than commercial liquid containing surfactants	Cunningham et al. (2004)	t3.8 t3.9 t3.10 t3.11
Vermiculite	PAH	<i>Bacillus</i> and <i>Mucor</i>	Best results than free culture	Dan et al. (2006)	t3.12 t3.13
Ca alginate	Cd	<i>Streptomyces</i> and <i>Bacillus</i>	Less effective than culture type	Je'ize'iquel and Lebeau (2008)	t3.14 t3.15
Alginate-lignin	Phenanthrene	<i>Phanerochaete chrysosporium</i>	Best results than free culture	Zhang et al. (2008)	t3.16 t3.17
Phosphorylated polyvinyl alcohol	Atrazine	<i>Agrobacterium radiobacter</i> and mix culture	Best results than free culture	Siripattanakul et al. (2008)	t3.18 t3.19 t3.20
Alginate and biofilm on tezontle	Organophosphate pesticides (OP)	Bacterial consortium OP adopted	Best than suspension culture	Yañez-Ocampo et al. (2009)	t3.21 t3.22 t3.23
Ca alginate	Diuron herbicide	<i>Delftia acidovorans</i> and <i>Arthrobacter</i>	One immobilized strain has better result than free culture	Bazot and Lebeau (2009)	t3.24 t3.25 t3.26 t3.27
Sodium alginate and diatomite	TPH	Hydrocarbon degrader mix consortium	Less effective than free inoculum and control plant	Farhana et al. (2012)	t3.28 t3.29 t3.30 t3.31

produced by plant roots. It inferred that the degradation potential is associated with functional changes in microbial community. 179  
180

Rhizospheric biodegradation is the smart technique which utilizes sunlight through plant to degrade pollutants with indigenous microbes. Due to limited methodological capabilities, there are many challenges which are difficult to explore; these includes which mechanisms influence microbial community composition in the rhizosphere, its type, degradative genes involved, and the effect of bioaugmentation in the rhizosphere. In order to assess the influence of plants on pollutant degradation, appropriate experimental design is an essential part of research study so that the conclusive results of pot experiments can be applied in the field plots or in scientific field studies (Mackova et al. 2006). 181  
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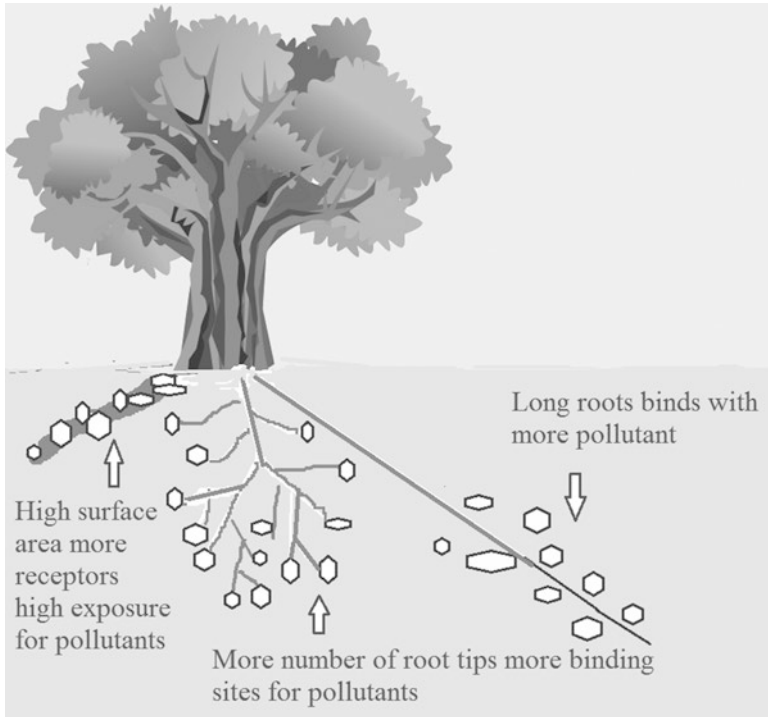
**Fig. 7.1** Bacterial community attaches with rhizosphere due to mucilage, and negative interaction of rhizospheric and inoculated bacteria takes place due to competition for nutrients and niche

#### 190 7.2.4 Plant Species Selection Criteria for Rhizodegradation

191 Plant root growth parameters are particularly important in the petroleum degradation.  
 192 The large 0 roots penetrate deeply in the impermeable soil layers, and its root-associated  
 193 microbial flora is exposed to more contaminants; similarly higher number of root  
 194 tips provides more binding site for contaminants (Fig. 7.2). The monocot plant species  
 195 have great potential of contaminant degradation due to its dense root systems which  
 196 provide a high surface area to soil-microorganism interaction for biodegradation  
 197 (Hussein 2006). In order to improve the efficiency of rhizospheric pollutant degradation,  
 198 bacterial inoculation on plant's seed could be an important strategy (Kuiper  
 199 et al. 2004).

200 The successful study of rhizodegradation of petroleum hydrocarbon was  
 201 achieved with *Sesbania cannabina* plant; this plant tolerates anoxia in the root  
 202 zone which is a prevailing condition of petroleum-polluted soil.

203 Observation of natural revegetation at the site can provide additional informa-  
 204 tion on potential plant species. The use of native versus non-native plants for the  
 205 rhizoremediation of contaminated sites is an important point of concern. In most  
 206 situations, plants that are native to the region of contamination have shown to be  
 207 most appropriate for rhizoremediation (Merkl et al. 2004). Species chosen for  
 208 rhizoremediation should have good adaptability to new environment and climatic  
 209 conditions of the region. This means that average temperature, annual rainfall,  
 210 and length of growing season are important considerations in rhizoremediation



**Fig. 7.2** Effect of root growth in petroleum rhizodegradation, morphology of roots plays an important role in binding of pollutants with roots which ultimately increases the pollutant degradation potential either by co-metabolic process or by pollutant uptake

planning (Frick et al. 1999) and each country have to recognize indigenous plants that can be utilized for phytoremediation (Robson et al. 2003). The introduction of non-native plants into any agricultural ecosystem is not always possible, and practical considerations such as cost and availability of seed are also very important.

### 7.3 Conclusion

In the bioremediation breakdown of the crude oil contaminant due to microbial activity enhanced in the presence of plant rhizosphere, the selection of native dominant plant species is required which can tolerate the exposed crude oil concentration. Bioaugmentation by using autochthonous microbes is of great interest because it is found to be best adopted with the contaminated environment. Inoculum addition or biostimulation techniques appeared to be effective in enhancing biodegradation of oil hydrocarbons in soil when used alone. However, in the presence of plant, bioaugmentation with free and sodium alginate-immobi-

225 lized cultures was ineffective as sodium alginate degrades very quickly and plant  
 226 growth required long time. One other reason was presence of root exudates which  
 227 select bacterial species and their subsequent activity. Natural plant-microbe inter-  
 228 action seems to be responsible for higher hydrocarbon degradation directly and  
 229 indirectly by promoting plant growth. The reason for ineffectiveness of bioaug-  
 230 mentation was that the inoculated culture was isolated from the freshly contami-  
 231 nated site while the soil sample was composted and weathered having more  
 232 recalcitrant fraction, so the stage of degradation was different which required dif-  
 233 ferent microbial community with relevant substrate-utilizing pathway which can  
 234 degrade petroleum hydrocarbon in a better way.

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# Author Queries

Chapter No.: 7 483176\_1\_En\_7\_Chapter

Queries	Details Required	Author's Response
AU1	Please check and confirm if the author names and affiliations are presented correctly.	
AU2	Please check if edit to sentence starting "Sometimes biodegradation..." is okay.	
AU3	Please check if edit to sentence starting "The second method..." is okay.	
AU4	Please check if edit to sentence starting "Among these autochthonous..." is okay.	
AU5	Please check whether the presentation of legend is fine for Table 7.2.	
AU6	Please check if edit to sentence starting "It is highly..." is okay.	
AU7	Please check if edit to sentence starting "Examples of analogy..." is okay.	
AU8	Please note that citation of references "Zhang et al. 2010; Gao et al. 2011; Kästner and Miltner 2016" are cited in the body but its bibliographic information are missing. Kindly provide its bibliographic information. Otherwise, please delete it from the text/body.	
AU9	References "Atlas (1991), Cassidy et al. (1996), Lee et al. (2008), Lu et al. (2010), Prince (1992), Suthersan (1999), Vogel (1996), Wiesel et al. (1993)" were not cited anywhere in the text. Please provide in text citation or delete the reference from the reference list.	